TABLE I

COMPARISON OF O¹⁸ IN 4,5-DIMETHYLCATECHOL FORMED Enzymically from 3,4-Dimethylphenol^a in $O^{18}_2^b$ and H₂O AND IN O₂ AND H₂O^{18°}

1120 AND IN 02 MID 1120			
Experiment	Found	Atom % excess O ^{18d} theoretical for uptake of one atom	No uptake
$O^{18}_2 + H_2O$	0.52	0.59	0.00
	.51		
	, 56		
$\mathrm{O}_2 + \mathrm{H}_2\mathrm{O}^{18}$.00	0.59	0.00
	.00		

^a Twenty-five ml. reaction volumes contained 0.3 minole ascorbic acid, 1.3 mmole KH_2PO_4 , 2.15 mmole K_2HPO_4 , 0.45 mmole 3,4-dimethylphenol and 4.0 mg. purified⁵ mushroom phenolase having 20-80 cresolase⁶ and *ca*. 1000 catecholase⁷ units/mg. dry wt. 4,5-Dimethylcatechol (30-50% yield) was isolated through its lead salt, from an ether extract of the reaction mixture, m.p. 84-86°. No hydroxylation occurred in the system when heat-denatured enzyme was sub-stituted for active protein. ^b Prepared electrolytically. ^c Obtained from the Stuart Oxygen Company, containing 1.4 atom % O¹⁸. ^d Mass spectrometry was performed by the Consolidated Engineering Corporation on carbon dioxide samples obtained by Unterzaucher pyrolysis⁸ of 4,5-dimeth-ylcatechol samples. Oxygen recovery was quantitative.

Since the phenolase complex is a cuprous protein^{9,10,11,12} which is in the cupric form after each hydroxylation¹³ and which combines with inhibitor

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(13) Hydroxylation does not proceed in the absence of reducing agents: cf. R. C. Behm and J. M. Nelson, THIS JOURNAL, 66, 711 (1944); M. Suda, N. Kimoto and S. Naono, J. Biochem. Soc. (Japan),

26, 603 (1954); A. B. Lerner, T. B. Fitzpatrick, E. Calkins and W. H.

CO in the ratio 2 Cu⁺/CO,⁹ hydroxylation by this enzyme system is describable as

(1)
$$\operatorname{Protein} \underbrace{\overset{\operatorname{Cu}^{+}}{\overset{\operatorname{Cu}^{+}}{\operatorname{Cu}^{+}}}_{\operatorname{Cu}^{+}} + O_{2} \longrightarrow \operatorname{Protein} \underbrace{\overset{\operatorname{Cu}^{-}}{\overset{\operatorname{Cu}^{-}}{\operatorname{Cu}^{-}}}_{\operatorname{Cu}^{-}}O$$
(2)
$$\operatorname{Protein} \underbrace{\overset{\operatorname{Cu}^{+}}{\overset{\operatorname{Cu}^{+}}{\operatorname{Cu}^{+}}}}_{\operatorname{Protein} \underbrace{\overset{\operatorname{Cu}^{+}}{\overset{\operatorname{Cu}^{+}}{\operatorname{Cu}^{+}}}}_{\operatorname{Cu}^{+}} + \operatorname{Diphenol} + H_{2}O$$
(3)
$$\operatorname{Protein} \underbrace{\overset{\operatorname{Cu}^{+}}{\overset{\operatorname{Cu}^{+}}{\operatorname{Cu}^{+}}}}_{\operatorname{Cu}^{+}} + 2e^{-} \longrightarrow \operatorname{Protein} \underbrace{\overset{\operatorname{Cu}^{+}}{\overset{\operatorname{Cu}^{+}}{\operatorname{Cu}^{+}}}}_{\operatorname{Cu}^{+}}$$

The hydroxylative function of phenolase (eq. 1 and 2) is thus coupled to an electron source (eq. 3), *i.e.*, oxidation of o-diphenol to o-quinone, which may be linked in turn to the common pathways of metabolism through TPNH+14 or DPNH+15, possibly by quinone reductase.¹⁶ The function of the phenolase complex as a terminal oxidase will be in demand during the biosynthesis of o-diphenols from monophenols. We propose that these o-diphenols are subsequently utilized to form flavonoids, lignins, tannins, cuticulation diphenols of arthropods, melanoproteins of chordates, and possibly adrenaline and noradrenaline.12 Some instances of light-irreversible inhibition of terminal respiration by carbon monoxide17,18 may be accounted for in these terms.

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BOOK REVIEWS

Annual Review of Physical Chemistry. Volume 5. G. K. ROLLEFSON, Editor, University of California, and R. E. POWELL, Associate Editor, University of California. Annual Reviews, Inc., Stanford, California. 1954. ix + 540 pp. 16 × 23 cm. Price, \$7.00.

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